

Chemical Papers

Surface modification and characterization of nanocellulose derived from the leaves of *Borassus flabellifer* --Manuscript Draft--

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Full Title:	Surface modification and characterization of nanocellulose derived from the leaves of <i>Borassus flabellifer</i>
Article Type:	Short Communication
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3 **Surface modification and characterization of nanocellulose derived from**
4 **the leaves of *Borassus flabellifer***

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Abstract

Nanocellulose (NC) is revolutionizing the world of biopolymers. This research explores the potential of NC as an additive for polymer composites, focusing on enhancing their structural and physio-chemical properties. The study involves the extraction of NC from *Borassus flabellifer* leaves and surface modification using lactic acid (LA). The composition of the leaves was estimated to be 62.2% cellulose, 15.8% hemicellulose, and 12.2% lignin by weight %. NC was extracted by subjecting pre-processed leaves to alkali treatment, resulting in an average particle size of 317 nm. NC and surface-modified NC (SMNC) were characterized using various bioanalytical techniques, including SEM, FTIR, and XRD, to evaluate the effectiveness of the surface modification. The water absorption test was performed to assess the surface modification of NC. The results confirmed the successful surface modification of NC using LA, as evidenced by the presence of LA functional groups on the NC after modification. XRD analysis confirmed the crystalline nature of NC and SMNC, with a higher crystalline index of 48.27% for SMNC compared to 42.78% for NC. The water absorption activity of SMNC was also measured and found to be lower than that of NC, suggesting that SMNC may be a promising additive for polymer composites. Overall, this study demonstrates the potential of SMNC for various applications in materials science.

Keywords *Borassus Flabellifer*. Nanocellulose. Surface modification. Lactic Acid

Introduction

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4 Natural fibers are a renewable and sustainable source of biodegradable materials that
5 offer several advantages over synthetic fibers. They are lightweight, low cost and widely
6 available, making them an attractive option for use as reinforcements in polymeric matrices.
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8 By extracting natural fibers from various plant parts, such as the bark, stem, root, fruits, leaves
9 and flowers, new bio-based composites can be created with enhanced properties. *Borassus*
10 *flabellifer* is among the sources of natural fiber that can be used as biodegradable materials [1].
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12 *Borassus flabellifer*, a member of the Arecaceae family, is native to Africa but is primarily
13 grown in Indo-Malaysia and Australia. It is usually referred to as the Palmyra palm. *Borassus*
14 *flabellifer* leaves are used to make umbrellas, mats, hats, fans, and writing material;
15 nevertheless, different parts of the plant have diverse scientific qualities that may be utilised in
16 different dimensions of research. For example, cellulose from the leaves has many applications
17 in the food packaging industry [2]. Cellulose, a polysaccharide composed of linear D-glucose
18 chains linked together by β -1,4-glycosidic bonds, is one of the significant components of the
19 leaves of *Borassus flabellifer*. Natural fibers, which include cellulose, hemicellulose, and
20 lignin, are commonly utilized by biopolymers to improve their physicochemical properties [3].
21 Among these, cellulose is the most abundant and primarily produced by plants. Natural fibres
22 can be purified for biopolymer applications through alkali treatment, which selectively
23 removes impurities such as hemicellulose and lignin, leaving behind pure cellulose. This
24 treatment effectively reduces the presence of hydrophilic components on the surface of the
25 fiber, increasing cellulose content [4].
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52 One of the unique substances derived from cellulose is NC, which is obtained through
53 various extraction methods. Due to its greater surface area, aspect ratio, and Young's modulus,
54 NC has an advantage over ordinary cellulose fibers [5]. The properties of NC are determined
55 by the source, the isolation method, and subsequent surface modifications [6]. NC is reinforced
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1 into various biopolymers to improve its physio-chemical properties [7,8]. The properties such
2 as biocompatibility, biodegradability, and adaptable surface chemistry make it suitable for
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4 biopolymer reinforcement [9]. The NC added as an additive is thought to improve the strength
5 and stiffness of the biocomposite formed, thereby improving the probability of its industrial
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7 use [10].
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13 Moreover, being biodegradable, NC is considered safe for use. However, unmodified
14 NC has certain disadvantages, making it less suitable for some applications. One of the major
15 drawbacks of unmodified NC is its hydrophilic nature. Hence, it has a strong tendency to absorb
16 moisture. This high water absorption rate can reduce the mechanical properties of materials
17 made from NC, making it less suitable for applications where moisture resistance is crucial,
18 such as in packaging materials. The high water absorption rate can also lead to swelling, which
19 can cause dimensional changes in the material, affecting its overall stability and durability.
20 Furthermore, due to its hydrophilic nature, NC has poor compatibility with hydrophobic
21 polymers, such as polylactic acid (PLA) [11]. This lack of compatibility can result in phase
22 separation and reduced interfacial adhesion between NC and the hydrophobic polymer matrix.
23 This can lead to decreased mechanical strength and stiffness of the resulting composite
24 material. Another disadvantage is its tendency to form aggregates due to its high surface area
25 and reactivity, which can lead to difficulties in processing and handling [12]. These aggregates
26 can also cause non-uniform dispersion of NC in the polymer matrix, which can further affect
27 the mechanical and physical properties of the resulting material. Surface modifications are,
28 therefore, performed to overcome these disadvantages. Specifically to reduce the
29 hydrophilicity of NC [13]. These modifications can improve its compatibility with polymer,
30 improving the resulting nanocomposites' mechanical, thermal, and barrier properties.
31 Introducing ester groups can also improve the dispersion of NC in polymer, reducing the
32 aggregation and improving the interfacial adhesion between the two materials. This can lead
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1 to more sustainable and eco-friendly materials with improved properties, making them suitable
2 for various applications, such as packaging, biomedical devices, and drug delivery systems.
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5 Although numerous researchers have documented the production of NC from *Borassus*
6 *flabellifer* as an additive. A very limited study has been conducted on surface modification of
7 NC extracted from the leaves of *Borassus flabellifer* using LA and its use as an additive for
8 polymer composite. This study aims to synthesise the NC from the leaves of *Borassus*
9 *flabellifer* and surface modify it with LA [14] so that it can be used as an additive for enhancing
10 the mechanical, physio-chemical, and gas barrier properties of biopolymer and its widespread
11 applications.
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22 **Experimental**

23 **Materials**

24 HCl, NaOH, H₂O₂, LA and SnCl₂ were used as a commercial-grade reagent from Sisco
25 Research Laboratories Pvt. Ltd, India. *Borassus flabellifer* leaves were gathered from Potheri,
26 Chennai, India. NC was extracted from these leaves, and it was subsequently modified to
27 SMNC.
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41 **Proximate composition of *Borassus flabellifer* leaves**

42 Cellulose, hemicellulose, and lignin content were estimated for *Borassus flabellifer*
43 leaves. The chemical compositions were estimated using gravimetric analysis. Equation (1)
44 was used to calculate the chemical composition of cellulose [15], hemicellulose [16] and lignin
45 [1].
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$$55 \text{ } Wt (\%) = \frac{(H1-H2)}{H1} \times 100 \dots\dots\dots(1)$$

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H1 is the weight of the untreated leave, and H2 is the weight of the treated leaves.

Pre-processing and pre-treatment of *Borassus flabellifer* leaves

The leaves samples were collected and processed further. The pre-processing of leaves was performed by mechanical operation technique known as ball milling. The obtained powder was alkali treated with 4% NaOH (1M) solution at a ratio of 1:10 and incubated at 80 °C for 3 h, followed by filtration. The alkali-treated biomass was bleached with 3% H₂O₂ and 4% NaOH (1M). The sample was again incubated at 80 °C for 3 h, followed by filtration and collection of the biomass. The biomass was ultrasonicated for 30 min at 20 kHz with a power input of 750 W, with an amplitude of 30% and pulses of 10 seconds on and off [17]. Subsequently, the samples were kept, and NC extraction was carried out.

Synthesis of NC

The ultrasonicated sample was centrifuged for 12 min at 10,000 rpm, and the supernatant was collected. The collected supernatant was lyophilized for 4 h for converting into powder form.

Surface modification of NC with LA

In the presence of the SnCl₂ (100 µg), NC aqueous solution was blended with LA at a ratio of 15:1. The sample was homogenized at 5000 rpm for 10 min, followed by ultrasonication for 1 h at 20% amplitude with a pulse of 10 sec on and off. The processed NC was kept in an 80°C water bath for 10 h. The sample was then filtered and air-dried [18].

Characterization studies of NC and SMNC

The extracted NC was characterized with a zeta sizer to confirm the nanoform. To confirm the particle size, the extracted NC was examined at 516 nm, 517 nm, and 518 nm

wavelengths [19]. The untreated leaf, untreated leaf with mechanical operation, alkali-treated leaf, NC and SMNC were then analyzed using SEM at 40,000 X magnification to determine surface morphology [20]. Following that, NC and SMNC were analyzed by FTIR and XRD. FTIR findings revealed the existence of functional groups. KBr pellets were used to prepare the samples with 10 mm in diameter and 1 mm in thickness for FTIR analysis. 5 mg of NC and SMNC were analysed by Agilent Technologies, Cary 600 Series / a Perkin-Elmer infrared spectrophotometer between the frequency range of 400–4000 cm⁻¹. The crystalline/amorphous nature of the particles and the crystallinity index were determined by performing XRD analysis at room temperature with a wavelength of 1.54060 Å with a Cu-Kα source and a generator at 40 kV 15 mA [21]. The crystallinity index (CrI) was calculated using the diffraction intensities of the crystalline structure and the amorphous fraction. Eqn. 2 is used to calculate the crystallinity index,

$$\text{CrI}\% = \frac{[I_{002} - I_{\text{am}}]}{I_{002}} * 100 \dots\dots\dots(2)$$

Where,

I₀₀₂ is the maximum intensity of the diffraction peak, and I_{am} is the intensity of the amorphous diffraction peak [22].

Water absorption test of NC and SMNC

The water absorption test was conducted using the gravimetric method. 1 g of NC and SMNC was taken, and their dry weight was recorded. Both samples were dipped in 50 mL of water. Furthermore, the weight change was recorded every 5 min until it obtained a constant weight. Water absorption was determined by measuring the increase in mass as a percentage of dry mass, and the moisture content value was calculated [23].

Results and Discussions

Proximate composition of *Borassus flabellifer* leaves

The chemical composition of cellulose, hemicellulose and lignin in untreated *Borassus flabellifer* leaves was evaluated using the gravimetric approach. Estimates showed that the leaves contain 64.2% cellulose, 15.8 % hemicellulose, and 12.2% lignin. Similar findings were reported by Singh et al. for untreated *Borassus flabellifer* leaves, with 68.1% cellulose, 14.5% hemicellulose, and 11.5% lignin contents [1].

Zetasizer Analysis

The Zetasizer is a powerful analytical instrument used to measure the size of particles in solution. The instrument utilizes dynamic light scattering (DLS) to determine particle size. To validate the isolated NC's particle size, zeta analysis was performed. The extracted NC's particle size distribution is depicted in Fig. 1. The NC isolated from leaves of *Borassus flabellifer* has particles with a diameter of 317 nm. Phanthong et al. [24] demonstrated that the particle size of NC ranges from 100 nm to 1000 nm, and a similar type of result was obtained for NC isolated from the leaves of *Borassus flabellifer*.

SEM Analysis

Scanning Electron Microscopy (SEM) is a powerful analytical technique used to investigate samples' surface morphology and composition. SEM analysis was performed to analyze the surface morphology of the untreated leaf, untreated leaf with mechanical operation, alkali-treated leaf, extracted NC and SMNC. The images were recorded at 40,000 X (Fig. 2). The SEM image recorded that the untreated leaf has a flat and sleek surface, untreated leaf with mechanical operation has large particles, while damaged and clump types particles were ascertained for alkali-treated leaf. An analogous kind of picture was observed by Arasu et al.,

1 following a sleek surface for untreated leaves and a rough surface for treated leaves [25]. On
2 the other hand, the SEM images depicted that NC has a smooth and shiny surface for small
3 particles, whereas SMNC particles have a clump-like pattern with a rough surface. The rough
4 surface with a clump formation indicates that LA has bound with NC. According to Wei et al.,
5 surface modification of NC affects the degree of smoothness of the NC, resulting in a rough
6 surface with a clump-like development. When NC was modified with LA, a similar configuration
7 was detected. It suggests that the surface modification has caused changes at the nanoscale
8 level, which may have implications for the material properties and potential applications of NC
9 [18].

22 **FTIR Analysis**

26 Fourier Transform Infrared Spectroscopy (FTIR) is a technique used to identify and
27 analyze the chemical composition of a sample. In FTIR analysis, a beam of infrared light is
28 directed through the sample, and the energy absorbed by the sample is measured. The resulting
29 spectrum provides information about the functional groups present in the sample. FTIR spectra
30 were used to investigate the functional groups present in the NC, LA, and SMNC. The effect
31 of esterification for NC and SMNC through LA is shown in Fig. 3. The FTIR spectra show a
32 prominent peak for NC between 3000 and 3500 cm^{-1} and at 1637.53 cm^{-1} , confirming the
33 presence of functional groups in NC. The peak between 3000 and 3500 cm^{-1} presents evidence
34 for the OH stretching of the free OH groups on the cellulose, and the peak at 1637.53 cm^{-1}
35 represents the C-O stretching of the NC. The peaks obtained for LA were similar to those
36 observed by Devi et al. [26]. Followed by, FTIR analysis of SMNC was performed, which
37 revealed that the surface modification occurred due to a shift in the intensity between the
38 functional groups. The intensity of the OH group was seen to be reduced in SMNC compared
39 to that of NC. The acidic group reacted with the alcohol group, resulting in the esterification
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1 process. Moreover, a carbonyl group formation at 1720 cm^{-1} has been observed in the SMNC
2 with LA, demonstrating that the surface modification was effective from a chemical standpoint.
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5 **XRD Analysis**

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9 X-ray diffraction (XRD) analysis is a powerful technique used to determine a
10 crystalline material's atomic and molecular structure. XRD works by shining a beam of X-rays
11 at a sample and then measuring the angles and intensities of the scattered X-rays. This data can
12 be used to calculate the positions of the atoms in the sample and their arrangement within the
13 crystal lattice. XRD analysis of NC and SMNC revealed the crystalline and amorphous nature
14 of the particles. The XRD pattern revealed that SMNC had a better crystalline structure than
15 NC (Fig. 4). A notable peak of about 21.75° and 21.83° was observed for both NC and SMNC,
16 respectively, confirming the findings of the study by Arun et al. [17]. The notable sharp peak
17 in SMNC revealed the crystalline nature of the particle. While no such prominent peak was
18 observed for NC, except a small peak around 21.75° . Further, the crystallinity index (CrI) of
19 both NC and SMNC was evaluated as 42.78 % and 48.27 %, respectively. This was confirmed
20 by the results of Robles et al. [27], who observed the crystallinity index of SMNC to be 53 %.
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39 **Water absorption test**

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42 The water absorption test is a widely used method for measuring the porosity and
43 permeability of materials. The test involves immersing a sample of the material in water for a
44 specific period and then measuring the amount of water the material has absorbed. The water
45 absorption test is used to assess the durability and resistance of a material to weathering,
46 erosion, and other forms of deterioration. The water absorption of NC and SMNC was studied
47 and represented in Fig. 5. In the first 5 min, rapid water uptake was observed for both samples.
48 Afterwards, the water uptake was reduced in the next 20 mins, after which a constant weight
49 was observed between 30 min to 50 min. The NC sample showed higher water absorption as
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1 compared to the SMNC. The result showed a 30 - 32 % reduction in water absorption of SMNC
2 in comparison to that of NC. The observed reduction of water absorption for SMNC was
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4 comparable with the findings of Sethi et al. [28], who observed a 33 – 35 % reduction in water
5 absorption of LA-modified NC compared to NC. With this observation, it can be pointed out
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7 that the surface modification of NC was successful, as reduced water absorption is vital for
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9 packing material.
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15 **Conclusion**

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18 Natural fibers, including cellulose and its derivatives, have attracted considerable
19 attention as eco-friendly alternatives to synthetic materials, particularly in packaging
20 applications. As a renewable and abundant resource, cellulose-based materials offer several
21 advantages, including biodegradability, low toxicity, and high strength. Furthermore, the
22 development of NC has expanded the potential applications of cellulose-based materials by
23 providing enhanced properties such as high surface area, aspect ratio, and mechanical strength.
24
25 In this study, NC was synthesized via ultrasonication from pre-processed *Borassus flabellifer*
26 leaves, and its average size was determined to be 317 nm using a zeta sizer. The synthesized
27 NC was then surface-modified using lactic acid (LA) through esterification in the presence of
28 SnCl₂. The resulting modified NC (SMNC) was characterized using various analytical
29 techniques such as SEM, FTIR, and XRD. The SEM analysis revealed the binding of LA with
30 NC, while the FTIR spectra confirmed the successful surface functionalization of the
31 synthesized NC through the presence of ester carbonyl peaks. The XRD analysis indicated the
32 particle nature of NC and SMNC. Furthermore, the water absorption test revealed that NC had
33 a higher water absorption capacity than SMNC, suggesting that the esterification of NC
34 successfully reduced its water absorption, which is crucial for packaging materials. Overall, it
35 highlights the importance of natural fibers, cellulose, and NC in developing sustainable
36 materials for various applications, including packaging. Further research in this area has the
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1 potential to lead to the development of eco-friendly materials that can reduce the environmental
2 impact of packaging waste while maintaining the desired performance characteristics. The
3 findings indicate that the surface modification of NC through esterification provides an
4 ecologically friendly technique to improve its physio-chemical characteristics. SMNC can be
5 blended with biopolymers such as polylactic acid (PLA) to create sustainable packaging
6 materials for food and beverage items. This research contributes to developing eco-friendly
7 packaging materials, potentially reducing the environmental impact of packaging waste.
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12 and Technology, for supporting this review work.
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14

15 **Ethical Approval**

16 Not applicable
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18 **Consent to Participate**

19 The authors have agreed to participate in the publication of the paper
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22 **Consent to Publish**

23 All authors have agreed to publish the paper
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25

26 **Authors Contributions**

27 **Ankit Chakraborty**: Formal analysis of data, validation and preparation of original
28 draft; **Pradnya Ghalsasi**: Collection of resources like protocols, and other related research
29 papers and reviewing original draft; **Radha P**: Conceptualization, supervision, writeup revision
30 - review and editing.
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Competing Interests

The authors declare no conflict of interest.

Availability of data and materials

The data used to support the findings of this study are available from the corresponding author upon request

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Figure legends

Fig. 1. Zeta-sizer analysis of nanocellulose

Fig. 2. SEM analysis of (a) Untreated leaf, (b) Untreated leaf with a mechanical operation, (c) alkali-treated leaf, (d) nanocellulose, (e) Surface modified nanocellulose

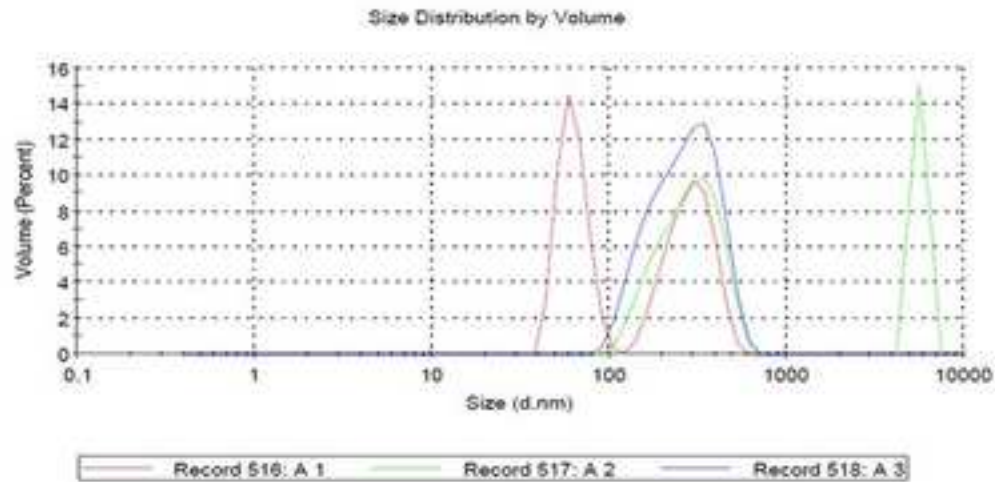
Fig. 3. FT-IR analysis of (a) nanocellulose, (b) lactic acid, (c) surface modified nanocellulose

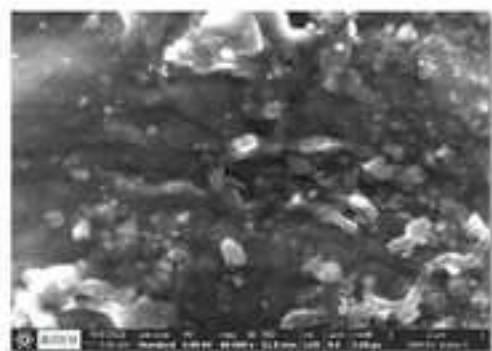
Fig. 4. XRD analysis of (a) nanocellulose (b) surface modified nanocellulose

Fig. 5. Water absorption test of (a) nanocellulose (b) surface-modified nanocellulose

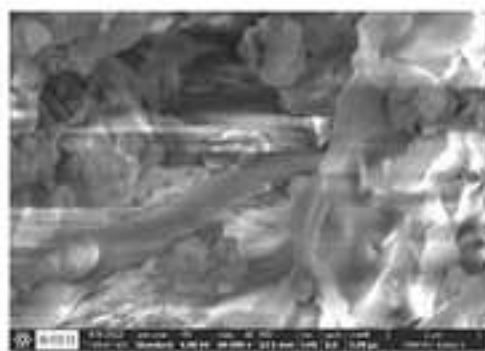
Results

	Size (d.nm):	% Volume:	St Dev (d.nm):
Z-Average (d.nm): 317.0	Peak 1: 292.5	51.9	83.11
Pdl: 0.490	Peak 2: 62.88	48.1	12.29
Intercept: 0.920	Peak 3: 0.000	0.0	0.000
Result quality Good			

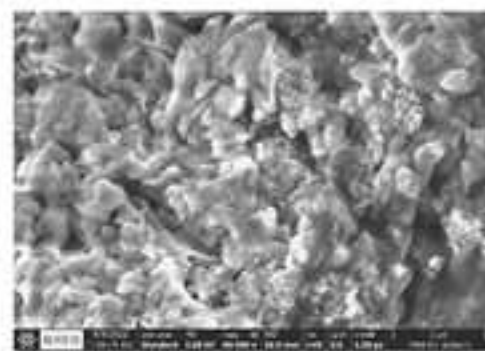




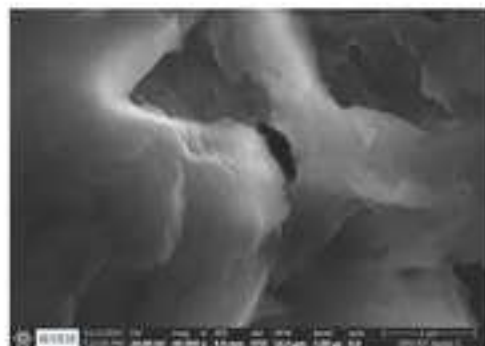
(a)



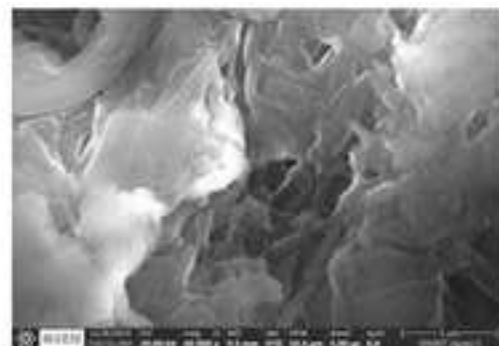
(b)



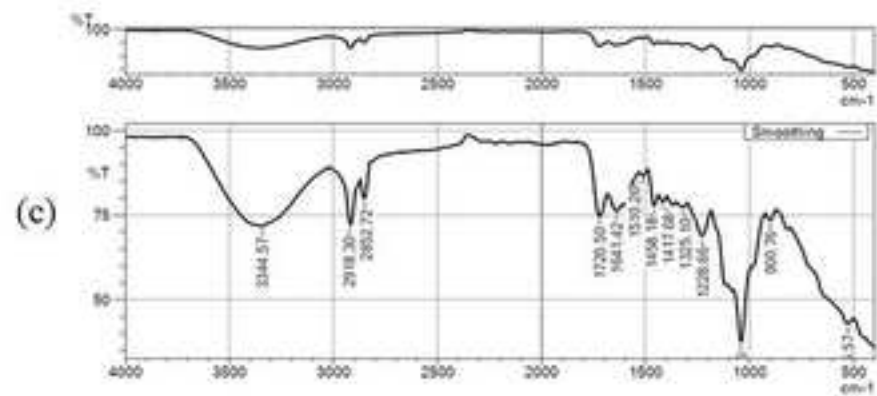
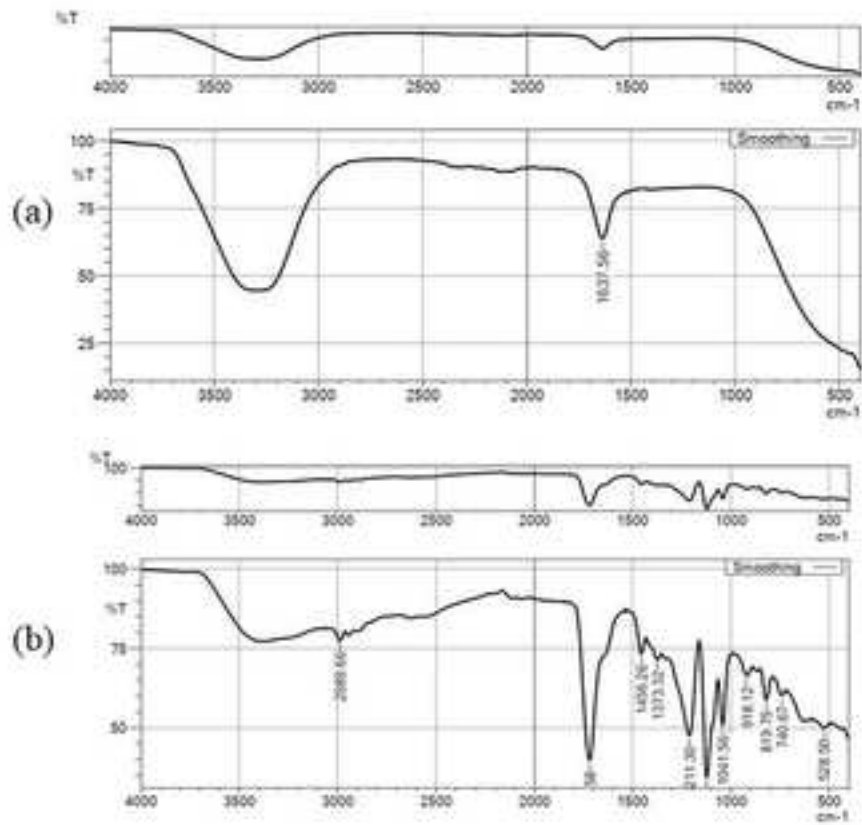
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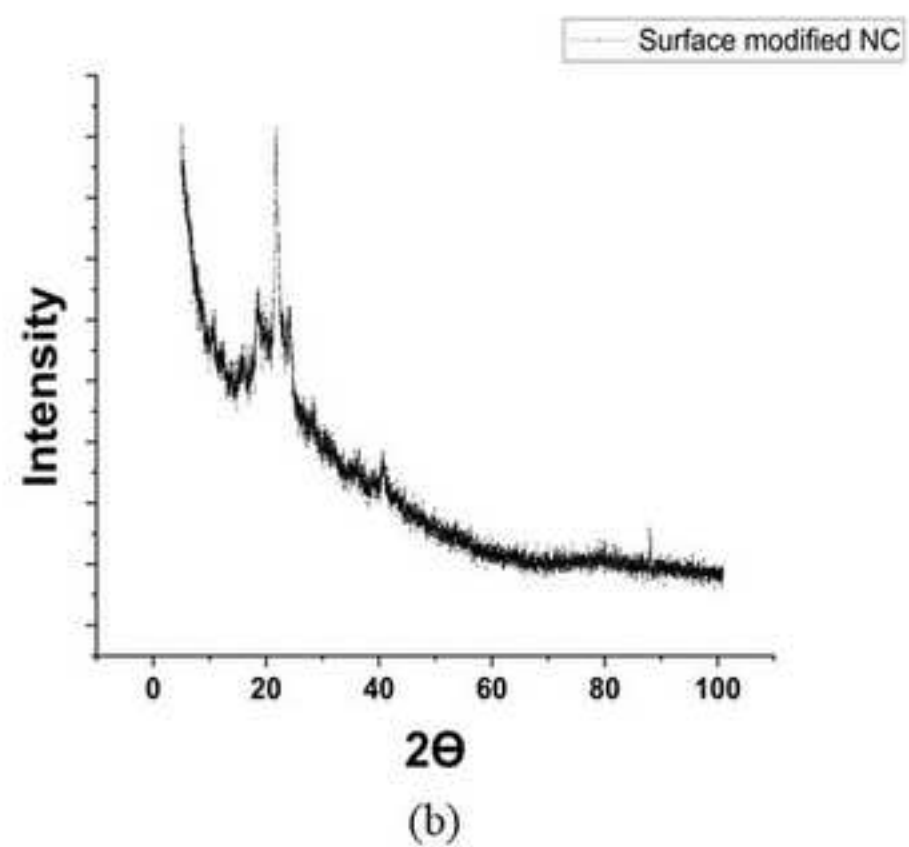
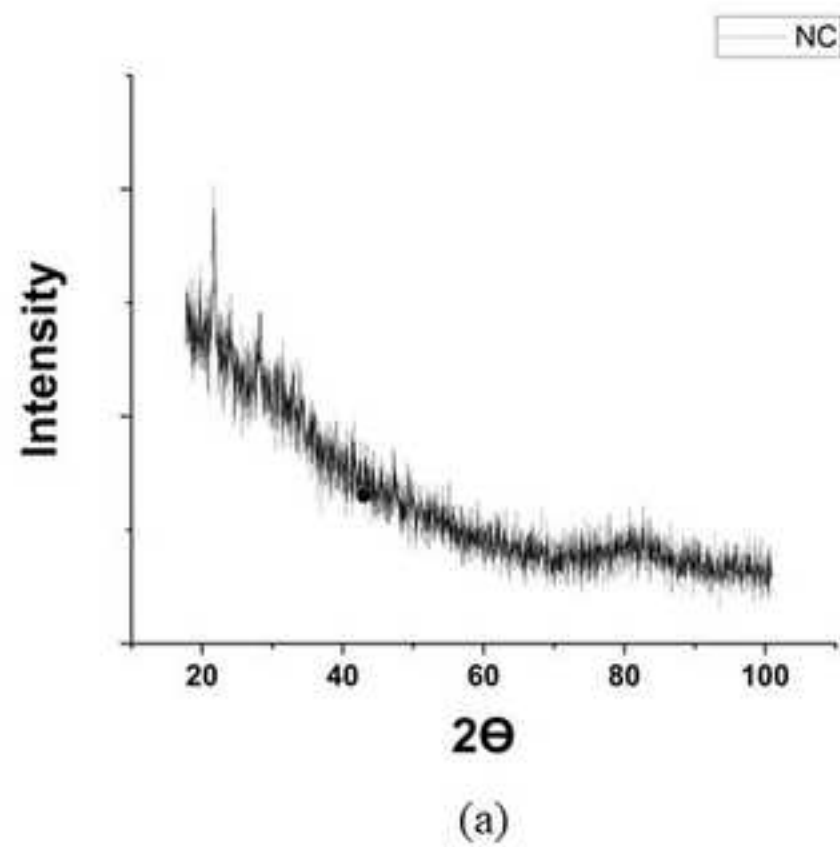


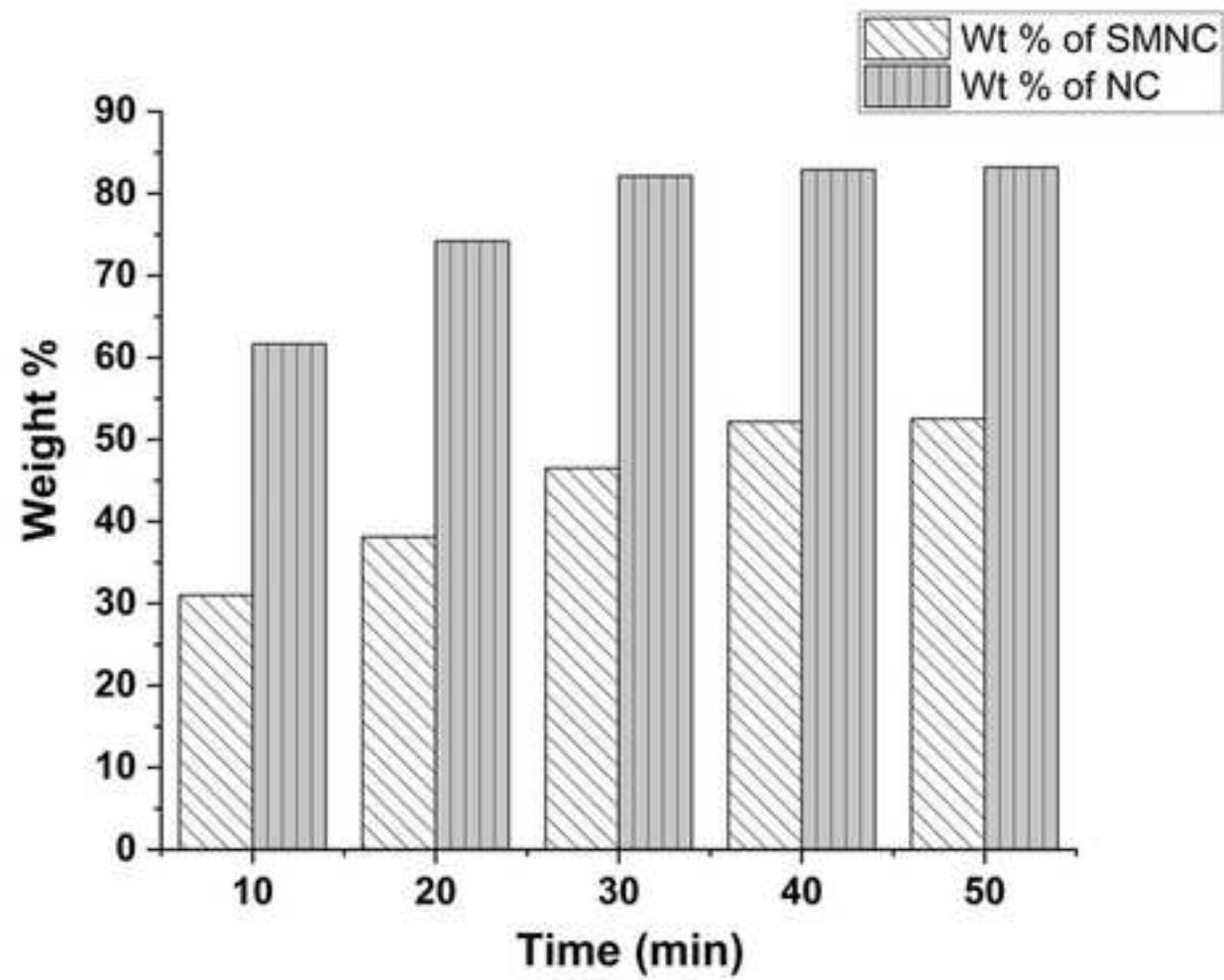
(e)



(f)









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