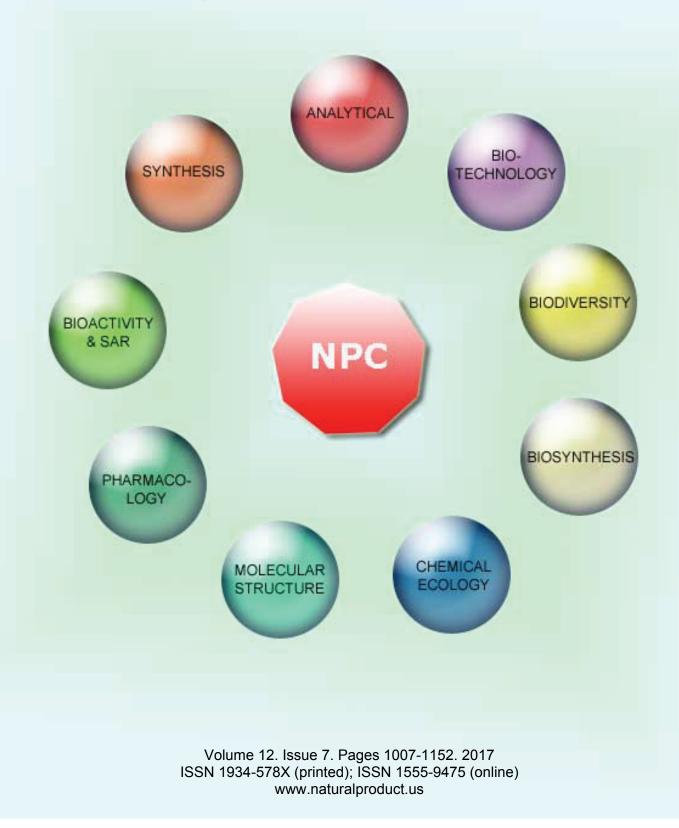
# NATURAL PRODUCT COMMUNICATIONS

### An International Journal for Communications and Reviews Covering all Aspects of Natural Products Research





# **Natural Product Communications**

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# NPC Natural Product Communications

### Phenolic Components from the Aerial Parts of Agrimonia pilosa

Nguyen Van Linh<sup>a</sup>, Hoang Le Tuan Anh<sup>b</sup>, Duong Thi Hai Yen<sup>b</sup>, Pham Thanh Ky<sup>a</sup>, Vu Manh Hung<sup>c</sup>, Nguyen Thi Thu Hien<sup>d</sup>, Trieu Quy Hung<sup>e</sup>, Nguyen Thi Cuc<sup>b</sup>, Duong Thi Dung<sup>b</sup>, Bui Huu Tai<sup>b</sup> and Phan Van Kiem<sup>b,\*</sup>

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Two new resorcinol derivatives, named agrimopilosides A and B (1 and 2), along with two known compounds, (2*S*, 3*S*)-aromadendrin 3-O- $\beta$ -D-glucopyranoside (3), (2*S*, 3*S*)-glucodistylin (4) were isolated from the aerial parts of *Agrimonia pilosa*. Their chemical structures were determined by mean of HRMS, NMR, CD spectra, and as well as by comparison with the reported data. At concentration of 20  $\mu$ M, compounds 1-4 modestly inhibited NO production in LPS-stimulated RAW264.7 cells, with the inhibitory rates in the range of 9.55-33.73%. None of them showed cytotoxicity toward HepG2, MCF-7, and SK-LU-1 human cancer cells by MTT assay.

Keywords: Agrimonia pilosa, Agrimopiloside A, Agrimopiloside B, Resorcinol derivative.

Agrimonia pilosa Ledeb. (Rosaceae) is widely distributed in southwest of Vietnam and has been used for treatment hemoptysis, hemorrhage, fever, diarrhea, and tuberculosis [1]. Previous phytochemical investigations of this genus confirmed the presence of flavonoids, triterpenoids, and phenolic compounds that possessed a variety of biological activities including anti-inflammatory, diabetes, anti-skin wrinkling, and whitening effects [2]. In the present study, we describe isolation and structural elucidation of two new phenolic glycosides 1 and 2 from the aerial parts of Agrimonia pilosa.

Compound 1 was obtained as a white amorphous powder. Its molecular formula was deduced to be C16H24O7 by HR-ESI-MS (m/z 327.1453 [M-H]), calcd for  $C_{16}H_{23}O_7$ , 327.1444), in conjunction with its <sup>13</sup>C-NMR data. The <sup>1</sup>H-NMR spectra of 1 showed three aromatic proton signals belonging to a 1,3,5trisubstituted benzene ring at  $\delta_{\rm H}$  6.45 (1H, t, J = 2.0 Hz), 6.41 (1H, t, J = 2.0 Hz), and 6.33 (1H, t, J = 2.0 Hz). The signal of an anomeric proton was observed at  $\delta_{\rm H}$  4.86 (1H, d, J = 7.5 Hz); two methyl groups were at  $\delta_{\rm H}$  1.20 (3H, d, J = 7.0 Hz) and 0.83 (3H, t, J = 7.0 Hz). The <sup>13</sup>C-NMR spectra of 1 revealed signals of 16 carbons which were classified into three non-protonated carbons, nine methines, two methylenes, and two methyl carbons. Of these, a set of carbinol carbon signals including  $\delta_C$  102.2, 78.1, 78.1, 74.9, 71.4, and 62.5 were typically characterized of a glucose unit. Shielded carbon signals ( $\delta_c$  43.2, 32.0, 22.3, 12.6) and their corresponding proton splitting pattern [ $\delta_H$  2.49 (1H, sex), 1.58 (2H, quin), 1.20 (3H, d), 0.83 (3H, t), J = 7.0 Hz each] suggested the presence of a sec-butyl group. Remain six aromatic carbon signals containing two deshielded carbons ( $\delta_{\rm C}$  160.1 and 159.3) indicated compound 1 to be a resorcinol derivative [3]. The HMBC correlations from protons H-10 ( $\delta_{\rm H}$  1.20) to carbons C-1 ( $\delta_{\rm C}$  151.4)/ C-7 ( $\delta_{\rm C}$  43.2)/ C-8 ( $\delta_{\rm C}$ 32.0); from H-7 ( $\delta_{\rm H}$  2.49) to C-1 ( $\delta_{\rm C}$  151.4)/ C-2 ( $\delta_{\rm C}$  107.9)/ C-6 ( $\delta_{\rm C}$ 109.2) confirmed the attachment of sec-butyl group to benzene ring. In addition, HMBC correlations from aromatic proton H-2 ( $\delta_{\rm H}$  6.45)

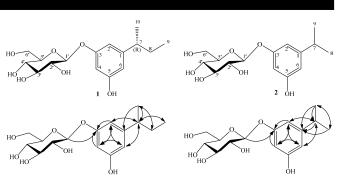


Figure 1: Chemical structures and key HMBC correlations of 1 and 2.

and anomeric proton H-1' ( $\delta_{\rm H}$  4.86) to carbon C-3 ( $\delta_{\rm C}$  160.1) were demonstrated the location of glucose moiety at C-3. The absolute configuration at C-7 of **1** was assigned to be *R* based on both negative CD bands at  $^{1}B_{\rm b}$  and  $^{1}L_{\rm a}$  transitions of benzene chromophore ( $\theta_{192}$  = - 1.84 and  $\theta_{228}$  = - 0.87 mdeg) and in comparison with the reported literature [4]. Consequently, chemical structure of compound **1** was established and named as agrimopilosides A.

Compound 2 was isolated as a white amorphous powder. The molecular formula of 2,  $C_{15}H_{22}O_7$ , was determined by pseudomolecular ion peak at m/z 313.1298 [M-H]<sup>-</sup> (calcd for  $C_{15}H_{21}O_7$ , 313.1287) in the HR-ESI-MS and <sup>13</sup>C-NMR analysis. The <sup>1</sup>H- and <sup>13</sup>C-NMR spectra of 2 (Table 1) were found to be similarity to those of 1, except signals of side-chain. Three shielded carbon signals and their corresponding proton splitting pattern [ $\delta_C/\delta_H$  35.5/2.79 (sep, J = 7.0 Hz), 24.3 and 24.2/1.21 (6H, d, J = 7.0 Hz)] indicated the presence of an isopropyl group in 2 instead of *sec*-butyl group in 1 [5]. Thus, compound 2 was established to be 1-isopropyl-3,5-dihydroxybenzene 3-*O*- $\beta$ -D-glucopyranoside, and named as agrimopiloside B.

Table 1: <sup>1</sup>H- and <sup>13</sup>C-NMR data for compounds 1 and 2.

D		$1^{a}$		<b>2</b> <sup>a</sup>
Pos.	$\delta_{C}$	$\delta_{\rm H}$ (mult., J in Hz)	δ <sub>C</sub>	$\delta_{\rm H}$ (mult., J in Hz)
1	151.4	-	152.6	-
2	107.9	6.45 (t, 2.0)	107.3	6.50 (t, 2.0)
3	160.1	-	160.1	-
4	102.5	6.41 (t, 2.0)	102.5	6.41 (t, 2.0)
5	159.3	-	159.3	-
6	109.2	6.33 (t, 2.0)	108.6	6.37 (t, 2.0)
7	43.2	2.49 (sex, 7.0)	35.5	2.79 (sep, 7.0)
8	32.0	1.58 (quin, 7.0)	24.2	1.21 (d, 7.0)
9	12.6	0.83 (t, 7.0)	24.3	1.21 (d, 7.0)
10	22.3	1.20 (d, 7.0)	-	-
1'	102.2	4.86 (d, 7.5)	102.2	4.86 (d, 7.5)
2'	74.9	3.45 (m)	74.9	3.45 (m)
3'	78.1	3.43 (m)	78.1	3.43 (m)
4'	71.4	3.42 (m)	71.4	3.42 (m)
5'	78.1	3.46 (m)	78.1	3.46 (m)
6'	62.5	3.72 (dd, 5.0, 12.0)	62.5	3.72 (dd, 5.0, 12.0)
		3.91 (dd, 2.0, 12.0)	62.5	3.91 (dd, 2.0, 12.0)

Measured in <sup>a</sup>CD<sub>3</sub>OD, Assignments were done by HSQC and HMBC experiments.

Remaining compounds were identified as (2S, 3S)-aromadendrin 3-O- $\beta$ -D-glucopyranoside (3) [6] and (2S, 3S)-glucodistylin (4) [7] on the basis of their NMR and CD spectral data which were in good agreement with those reported in the literature.

Compounds 1-4 were evaluated for their effects on NO production in the LPS-stimulated RAW 264.7 cells. The results showed that at concentration of 20  $\mu$ M, all compounds modestly inhibited NO production with the inhibitory rates from 9.55% to 33.73%. The isolated compounds 1-4 were also evaluated their effects on the proliferation of HepG2, MCF-7, and SK-LU-1 human cancer cells using MTT assay. However, none of them had significant cytotoxicity against the tested cell lines.

### Experimental

*General:* Optical rotations, Jasco DIP-370 automatic polarimeter; NMR, Bruker AM500 FT-NMR spectrometer; CD, Chirascan spectrometer; HR-ESI-MS, Agilent 6530 Accurate Mass Q-TOF LC/MS system.

**Plant material:** The aerial parts of *A. pilosa* were collected at Trung Khanh, Cao Bang province, Vietnam, in August, 2013. Its scientific name was identified by Dr. Pham Thanh Huyen, Institute of Ecology and Biological Resources, VAST. A voucher specimen (6695A) is deposited at the Herbarium of Military Institute of Traditional Medicine.

*Extraction and isolation:* The dried and powdered of aerial parts of *A. pilosa* (5.0 kg) were sonically extracted with methanol at 50  $^{\circ}$ C

### References

for three times (10.0 L each). After removal of the solvents, methanol extract (700.0 g) was suspended with distilled water (1.5 L) and successively partitioned with dichloromethane and ethyl acetate (three times, 2.0 L each) to give corresponding soluble extracts, dichloromethane (BAPD, 320.0 g), ethyl acetate (BAPE, 80.0 g), and water layer (BAPW). The BAPE extract (80.0 g) was separated on a silica gel column chromatography, eluting with gradient of dichloromethane/ methanol (100/0, 50/1, 30/1, 20/1, 10/1, 1/1, 0/100, v/v) to give seven fractions, BAP1A - BAP1G). Fraction BAP1D (10.0 g) was continuously separated on a RP-18 column chromatography, eluting with methanol/water (1/3, v/v) to give seven smaller fractions, BAP2A - BAP2G. Fraction BAP2C (0.5 g) was chromatographed on Sephadex LH20 column, eluting with methanol/water (1/1, v/v) to give compound 4 (30 mg). Fraction BAP2D (2.0 g) was separated on a silica gel column chromatography, eluting with dichloromethane/methanol/water (4/1/0.1, v/v/v) to give three fractions, BAP3A - BAP3C. Compound 3 (15 mg) was obtained from the BAP3B fraction (0.3 g) using a RP-18 column, eluted with acetone/water (2/5, v/v). Fraction BAP2E (1.0 g) was chromatographed on a RP-18 column, eluting with acetone/water (1/2, v/v) to yield three sub-fractions, BAP4A - BAP4C. Compound 1 (7.0 mg) was obtained from the BAP4A fraction (0.1 g) using a RP-18 column, eluted with acetone/water (1/2, v/v). Fraction BAP2F was chromatographed on a RP-18 column, eluting with acetone/water (1/3, v/v) to yield four sub-fractions (BAP5A-BAP5D). Compound 2 (15.0 mg) was obtained from the BAP5B fraction (0.2 g) using a Sephadex LH-20 column, eluted with methanol/water (1/1, v/v).

### Agrimopiloside A (1)

White amorphous powder.  $[\alpha]_{D}^{25}$ : +28.3 (*c* 0.1, MeOH). CD (MeOH):  $\theta_{192}$  = - 1.84 and  $\theta_{228}$  = - 0.87 mdeg <sup>1</sup>H-NMR (500 MHz, CD<sub>3</sub>OD) and <sup>13</sup>C-NMR (125 MHz, CD<sub>3</sub>OD): Table 1. HR-ESI-MS: *m/z* 327.1453 [M-H]<sup>-</sup>; calcd for C<sub>16</sub>H<sub>23</sub>O<sub>7</sub>, 327.1444.

### Agrimopiloside B (2)

White amorphous powder.  $[\alpha]_{D}^{25}$ : +41.6 (*c* 0.1, MeOH). <sup>1</sup>H-NMR (500 MHz, CD<sub>3</sub>OD) and <sup>13</sup>C-NMR (125 MHz, CD<sub>3</sub>OD): Table 1. HR-ESI-MS: *m/z* 313.1298 [M-H]<sup>-</sup>; calcd for C<sub>15</sub>H<sub>21</sub>O<sub>7</sub>, 313.1287.

**Supplementary data:** HR-ESI-MS, NMR, and CD spectra of compounds **1** - **4** can be found in the online version.

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